HUMAN ENDOGENOUS RETROVIRUSES AND IMMUNE TOLERANCE IN PREGNANCY

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BSc (Biotechnology) BSc (Biotechnology) (Hons Class 1)





Abstract

The human placenta expresses endogenous retroviral envelope proteins which have been postulated to play an important role in the physiology of pregnancy. Of these, syncytin-1 and syncytin-2 are highly expressed in the syncytiotrophoblast and cytotrophoblast respectively and are thought to be key factors in the regulation of syncytialisation due to their fusiogenic properties. In addition to their role in cell fusion, it has also been speculated that syncytin-1 and syncytin-2 may have a role in maternal immune tolerance due to the presence of the highly conserved immunosuppressive domain (ISD) within its sequence. However, no studies are yet to confirm this putative role. Another factor which has been speculated to have a role in maternal immune tolerance is Corticotropin Releasing Hormone (CRH) which has been shown to promote implantation and the maintenance of early pregnancy via the regulation of FasL. Interestingly, both syncytin-1 and FasL have been identified in immunosuppressive placental exosomes. Since CRH stimulates cyclic AMP (cAMP) production and syncytin-1, syncytin-2 and FasL are all stimulated by the cAMP second messenger pathway, it was hypothesised that syncytin-1 and syncytin-2 may be regulated by CRH. Further, it was hypothesised that syncytin-1 may contribute to the modulation of the maternal immune environment during pregnancy. To examine the regulation of syncytin-1 and syncytin-2 by CRH, a combined nucleic acid and protein extraction procedure was developed using column based nucleic acid extraction kits. Using 2D buffer, proteins extracted using this method were shown to have a comparable protein profile to conventionally extracted proteins. This method was then used to examine RNA and protein levels in CRH treated BeWo cells. Following CRH treatment of BeWo cells, a significant upregulation of syncytin-1, syncytin-2 and FasL mRNA was observed. CRH also increased the production of the syncytin-1 precursor in an exosomal fraction. To examine the immunosuppressive properties of syncytin-1, the recombinant ectodomains of human and mouse syncytins were produced and purified using a combination of affinity chromatography and gel filtration. The immunosuppressive properties of the syncytin-1 recombinant ectodomain were then tested using a whole blood culture model stimulated with LPS or PHA.

Syncytin-1 recombinant ectodomain at a concentration of 1μ M inhibited the production of TNF- α by 50% and CXCL10 by 65% in whole blood cultures following maximal stimulation with LPS. Syncytin-1 recombinant ectodomain also inhibited the production of IFN- γ by 30% in PHA stimulated PBMC. These studies demonstrate for the first time that syncytin-1 has immunosuppressive properties. Further, these studies show that CRH has a role in the stimulation of syncytin-1 and its subsequent sorting into exosomes. Circulating placental exosomes containing syncytin-1 and other immunosuppressive factors including FasL may interact with maternal immune cells to prevent an immune response against the fetal-placental unit. This is a novel mechanism that may contribute to our understanding of how a genetically different fetus can be tolerated by the mother during pregnancy.

Statement of Originality

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John Even Schjenken

July, 2011

Declaration

The work presented in chapter 2 was part of collaborations between John Schjenken and Jorge Tolosa. This work was also presented in the thesis of Jorge Tolosa as an appendice and has been published in the journal Biotechniques (1). http://dx.doi.org/10.2144/000112594

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- Figure 3.4 Representative silver stain and syncytin-1 western blot of exosomal proteins extracted from CRH treated BeWo cells. *A) Representative silver stain of CRH treated exosomal protein. B) Representative western blot of syncytin-1 protein expression in exosomes produced by BeWo cells after CRH stimulation. In these experiments, an equal number of BeWo cells were treated by CRH. The loading of these gels was therefore controlled by the number of BeWo cells treated with CRH. Observe the dose dependent increase in syncytin-1 protein in exosomes, reaching a significant peak at 50nM CRH treatment (lane 6). Results are representative of 3 experiments. M - Molecular Weight standard; Mark12 (Invitrogen); 1 – Vehicle for*

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Figure 4.7. Chromatogram of Ovalbumin and Ribonuclease A standards run using Superdex 75 10/300 GL column (GE healthcare) on ÄKTATM FPLCTM (GE healthcare). A) GE healthcare elution profile for 500µl standard (BSA (1) – 67kDa - 8mg/ml, Ovalbumin (2) – 43kDa – 2.5mg/ml, Ribonuclease A (3) – 13.7kDa – 5mg/ml, Apoprotinin (4) – 6.5kDa – 2mg/ml, Vitamin B12 (5) – 1.355kDa – 1mg/ml) after sample was run on Superdex 75 10/300 GL in 50mM phosphate buffer, 150mM NaCl, pH7 at 0.4ml/min. B) 500µl of 2.5mg/ml Ovalbumin (43kDa) and 5mg/ml Ribonuclease A (13.7kDa) was run through Superdex 75 10/300 GL in 25mM HEPES, 150mM NaCl, pH 8.5 at 0.15ml/min. Ovalbumin eluted in fraction 10l while Ribonuclease A eluted at 14ml.

Figure 4.10. Syncytin-A ectodomain purification. Syncytin-A ectodomain was induced in ImM IPTG and the insoluble inclusion body fraction was isolated using centrifugation methods. Syncytin-A ectodomain was initially purified by affinity chromatography using Hitrap IMAC HP methods (A, B). Complete purification was achieved through two runs through the Superdex 75 10/300 gel filtration column in 25mM HEPES, 150mM NaCl pH7.4 (C-F). A, C and E show a representative

Figure 4.11. Syncytin-B ectodomain purification. Syncytin-B ectodomain was induced in ImM IPTG and soluble bacterial protein was isolated using centrifugation methods. Syncytin-B ectodomain was then purified by affinity chromatography using Hitrap IMAC HP methods (A, B). Complete purification was achieved through two runs through the Superdex 75 10/300 gel filtration column in 25mM HEPES, 150mM NaCl pH7.4 (C-F). A, C and E show a representative graph/chromatogram of syncytin-B ectodomain following Hitrap IMAC HP methods (A) or Superdex 75 10/300GL purification (C and E – blue line). B, D and F show a representative Coomassie stained SDS-PAGE gel of syncytin-B ectodomain eluted from either Hitrap IMAC (B) or Superdex 75 10/300 GL (D or F) columns. Protein ladder = Mark12 (Invitrogen); Before (B) = Sample before application to Hitrap column; After = Sample after application to Hitrap column; #1-15 = Sample elution from column. 135

Figure 4.12. Representative Western blot of purified syncytin recombinant ectodomains. *Representative Western Blot of purified recombinant ectodomains of human syncytin-*1 and syncytin-2 (A-B and C-D); and mouse syncytin-A and syncytin-B (E-F and G-H) generated in E. coli. 2µg of purified protein was used. Layout of membranes are as follows; A) and B) 1 - Magic mark (Invitrogen), 2 – syncytin-1 ectodomain fraction 10, 3 – syncytin-1 ectodomain fraction 12; C) and D) 1 - Magic mark (Invitrogen), 2 – syncytin-2 ectodomain fraction 10, 3 – syncytin-2 ectodomain fraction 10, 3 – syncytin-2 ectodomain fraction 10, 3 – syncytin-A ectodomain fraction 10, 3 – syncytin-A ectodomain fraction 12; G and H 1 - Magic mark (Invitrogen), 2 – syncytin-B ectodomain fraction 10, 3 – syncytin-B ectodomain fraction 10, 3 – syncytin-B ectodomain fraction 10, 5 – syncytin-B ectodomain fraction 10, 6 – syncytin-B ectodomain fraction 10, 7 – syncytin-B ectodomain fraction 10, 7 – syncytin-B ectodomain fraction 12. A), C), E) and G) syncytin AbL Pre-Immune Sera (1:1000), B), D), F) and H) syncytin AbL Immune Sera (1:1000).

Figure 5.2. Syncytin-1 recombinant ectodomain elution profile. *A)* SDS-PAGE of purified syncytin-1 ectodomain. *B)* FPLC chromatogram of purified syncytin-1 ectodomain. Syncytin-1 recombinant ectodomain was produced in E.coli as a soluble His-tagged protein and purified using affinity chromatography (Hitrap) and gel filtration chromatography (Superdex 75 10/300GL). The recombinant protein eluted

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Figure 5.8. Inhibition of TNF-alpha production in whole blood stimulated with LPS using syncytin-1 recombinant protein from fraction 10-12. A dose dependent decrease in TNF-alpha production was observed following maximum stimulating doses of LPS (10ug/ml) and treatment with different concentrations of syncytin-1 recombinant protein. An inhibition of 45% TNF-alpha compared to the vehicle control was observed using 1uM syncytin-1 recombinant protein. Data are the mean from 5 individual experiments and the error bars refer to standard error. ** - p<0.01 compared to the no treatment control.
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recombinant protein was measured by the LDH assay. No significant difference in

LDH was observed between treatments and vehicles at any concentration of syncytin-1 recombinant protein. Data are the mean from 3 individual experiments and the Figure 5.10. Inhibition of CXCL10 production in LPS stimulated whole blood using syncytin-1 recombinant protein fraction 10. An inhibition of 65% CXCL10 compared to the vehicle control was observed using 1uM syncytin-1 recombinant protein following treatment of whole blood with maximal stimulating doses of LPS (10ug/ml). Data are the mean of 5 individual experiments and the error bars refer to standard Figure 6.1. Schematic of the possible role of syncytin-2 during pregnancy. *Recent* studies have suggested that syncytin-2 has immunosuppressive properties, however, the authors have not attempted to put this immunosuppressive role in context considering that syncytin-2 is expressed in the cytotrophoblast and is not in direct contact with the maternal circulation. This thesis proposes that the increasing CRH levels during gestation stimulate the production of syncytin-2 in the cytotrophoblast which interacts with the syncytin-2 receptor on the syncytiotrophoblast resulting in *cytotrophoblast cell fusion. STL* = *Syncytiotrophoblast layer, CT* = *Cytotrophoblast, S2R* = *Syncytin-2 receptor*, *CTDC* = *Cytotrophoblast Daughter Cell*, *CTSC* = Figure 6.2. Schematic of the possible immunosuppressive role of syncytin-1 throughout gestation. During pregnancy, the semi-allogenic fetus expresses antigens derived from both the mother and the father without immunological rejection. This thesis proposes that the increasing CRH levels during gestation stimulate the production of syncytin-1 and FasL which are sorted into exosomes along with a number of other immunosuppressive factors and released into the maternal circulation impairing the ability of circulating immune cells to mount an immune response against the semiallograft fetus resulting in maternal immune tolerance. STL = Syncytiotrophoblast *layer*, *CTL* = *Cytotrophoblast layer*.....181

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Abbreviations

CHAPS -	3-[(3-Cholamidopropyl)dimethylammonio]-1-propanesulfonate
ACTH -	Adrenocorticotropic hormone
AGRF -	Australian Genome Research Facility
bp -	Base pair
cDNA -	Complementary DNA
CRH -	Corticotropin Releasing Hormone
CRH-BP -	CRH binding protein
CRH-R -	CRH receptor
cAMP -	cyclic AMP
DISC -	Death Inducing Signalling Complex
DAF -	Decay Accelerating Factor
DTH -	Delayed type hypersensitivity
dH ₂ O -	Distilled water
ERV -	Endogenous retroviral/retrovirus
ERVs -	Endogenous retroviruses
env gene -	Envelope gene
ERK -	Extracellular signal-regulated kinase
EVT -	Extra villous trophoblast
FADD -	Fas-associated death domain
FasL -	Fas Ligand
FPLC -	Fast Protein Liquid Chromatography

FeLV -	Feline Leukaemia Virus
Gcma -	Glial cells missing a
GM-CSF -	Granulocyte macrophage colony stimulating factor
HBSS -	Hanks buffered salt solution
HPLC –	High Performance Liquid Chromatography
Hitrap IMAC -	Hitrap Immobilised Metal Affinity Chromatography
HRP -	Horse radish peroxidase
HERV -	Human Endogenous Retrovirus
HERVs -	Human Endogenous Retroviruses
HIV -	Human Immunodeficiency Virus
HLA -	Human Leukocyte Antigen
Ig -	Immunoglobulin
ISD -	Immunosuppressive domain
IDO -	Indoleamine 2,3,-dioxygenase
IFN -	Interferon
IL -	Interleukin
JAK3 -	Janus Kinase 3
IPTG -	Isopropyl β -D-1-thiogalactopyranoside
LDH -	Lactate Dehydrogenase
LAL -	Limulus Amebocyte Lysate
LPS -	Lipopolysaccharide
LCMS -	Liquid Chromatography mass spectrometry
LB –	Luria broth

MHC -	Major Histocompatibility Complex
MPMV -	Mason Phizer Monkey Virus \
MAC -	Membrane Attack Complex
MCP -	Membrane Co-factor Protein
mRNA -	Messenger RNA
μg -	Microgram
μΜ -	Micromolar
mg -	Milligram
mM -	Millimolar
MEK –	mitogen-activated protein kinase/ERK kinase
M -	Molar
MMulV -	Moloney Murine Leukaemia Virus
ng -	Nanogram
nm -	Nanometres
nM -	Nanomolar
NK cells -	Natural Killer cells
OD -	Optical density
1D -	One dimensional
PBMC -	Peripheral Blood Mononuclear Cells
PMA -	phorbol 12-myristate 13-acetate
PBS -	Phosphate buffered saline
Pi3-K -	phosphoinositide-3 kinase
PHA -	Phytohaemagglutinin

PCR -	Polymerase Chain Reaction
PIBF -	progesterone induced blocking factor
PD1 -	Programmed Death 1
PDL1 or PDL2 -	Programmed death Ligand 1 / 2
PKA -	Protein kinase A
RT-PCR -	Real-time Polymerase Chain Reaction
RCA -	Regulators of Complement Activation
T-reg cells -	Regulatory T cells
rpm -	Revolutions per minute
RT -	Room temperature
SDS-PAGE -	Sodium Dodecyl Sulphate Polyacrylamide Gel Electrophoresis
ASCT2 -	Solute Carrier Family 1 (neutral amino acid transporter),
member 5	
SEB -	Staphylococcal Enterotoxin B
SOB -	Super Optimal Broth
SOCS -	Suppressors of cytokine signaling
TB -	Transformation buffer
TGF -	Transforming Growth Factor
TAE buffer -	Tris-acetate-EDTA buffer
TBST -	Tris-buffered saline Tween 20
TE buffer -	Tris-EDTA buffer
TNF -	Tumour Necrosis Factor
2D -	Two dimensional

2D buffer -	Two dimensional electrophoresis lysis buffer
Th1 -	Type 1 helper T cells
Th2 -	Type 2 helper T cells
UV -	Ultra-violet